



An Update on Isolation of Extensively Drug Resistant (XDR) *Salmonella enterica* from Blood Cultures in a Tertiary Care Centre

Sadia Hameed, Mateen Izhar, Anwaar Basheer, Chetan Lal, Shazia Rishi, Abdul Basit
Department of Microbiology, Shaikh Zayed Medical Complex, Lahore

ABSTRACT

Introduction: Typhoid fever is a public health issue, burdening many regions of the world with poor socio-economic background. Management of this disease faces the major hurdle of antimicrobial resistance. The present study reveals current pattern of antimicrobial susceptibility among *Salmonella enterica* (*Salmonella typhi* & *Salmonella paratyphi* A) blood culture isolates from typhoid fever cases. Regular data collection about *Salmonella* infections and their response to antimicrobial agents, coupled with a long term commitment to providing adequate health information systems, is the key to effective planning and policy formation against typhoid fever. **Aims & Objectives:** To evaluate the prevalence of ceftriaxone resistant *Salmonella enterica* isolates from blood cultures in Shaikh Zayed Medical Complex for updating nosocomial antimicrobial resistance data. **Place and duration of study:** This research study was conducted at Shaikh Zayed Medical Complex, Lahore from March 2018 to May 2019. **Material & Methods:** Blood cultures positive for *Salmonella enterica* were analyzed, taking into consideration the gender & age of patients with typhoid fever. Antimicrobial susceptibility testing was carried out through disc diffusion method. The recommended antimicrobial agents for *S. typhi* (*Salmonella typhi*) & *S. paratyphi* A (*Salmonella Paratyphi* A) (as per CLSI, USA 2018/2019 guidelines; described later), were tested and analyzed during this study with the main focus on 3rd generation cephalosporin resistance pattern. Among second line treatment options, meropenem and azithromycin were selected for study. **Results:** Out of 899 isolates of *Salmonella enterica* (from 13964 samples for blood culture), 849 (94.4%) were *S. typhi* and 50 (5.6%) were *S. paratyphi* A. Of these, 57.1% isolates were from males, 42.9% from females, 81.7% from children (age less than 12 years) and 18.3% were from adults (age more than 12 years). A continuously rising resistance percentage was observed for 3rd generation cephalosporins over the span of fifteen months. It was 43.4% for ceftriaxone (CRO). No case of meropenem and azithromycin resistance was observed during study period. **Conclusion:** Demographic information was provided by this study regarding high level of resistance among *Salmonella enterica* isolates particularly *S. typhi*. The first line antibiotic drugs were a complete failure in ongoing outbreak of typhoid fever. The novel results of this study are the high resistance percentages for 3rd generation cephalosporins, whether oral or parenteral. This result is worrisome as it will leave us with no option but to resort to second line drugs. However, an increasing trend of blood culturing was observed in this research. Extensively drug resistant (XDR) *Salmonella* infection has definitely highlighted the importance of blood culture and its use as a more preferred diagnostic tool.

Key words: Typhoid fever, Extensively drug resistant (XDR) *Salmonella enterica*, (*S. typhi* & *S. paratyphi* A), Antimicrobial Resistance, CRO (Ceftriaxone) resistance.

INTRODUCTION

Six major disease outbreaks have been reported in Pakistan in year 2018: Extensively drug resistant (XDR) typhoid fever, chicken pox, dengue fever, poliomyelitis, chikungunya and crimean congo haemorrhagic fever (CCHF)¹. Typhoid fever is a systemic febrile illness requiring urgent antibiotic therapy. Approximately 12-27 million typhoid fever cases occur annually, worldwide². In 2014, out of

0.2 million deaths associated with typhoid fever, more than 90% occurred in Asia³.

Moreover, Pakistan and India account for a very high incidence of typhoid fever compared to other South Asian countries such as China, Indonesia and Vietnam⁴. The mortality rate rises up to 30% resulting from inappropriate treatment (as serious life threatening complications such as small bowel perforation and meningitis can occur), whereas it is reduced up to 0.5% with proper remedy^{5, 6}.

S. typhi accounts for a major count of typhoid fever cases. The relative contribution and incidence of *Salmonella paratyphi* A, a less severe disease causing agent than *S. typhi*, is not well understood, as most studies in Indian Subcontinent have mainly focused on *S. typhi*. However a rise in *S. paratyphi* A infection has been reported⁷.

Historically, the first line treatment options for typhoid fever have been chloramphenicol, ampicillin, and cotrimoxazole³. In early 1970s, resistance to chloramphenicol emerged in *Salmonella enterica* strains followed by resistance to ampicillin and cotrimoxazole. Since 1990s, many epidemics of typhoid fever occurred due to multidrug resistant *Salmonella typhi* (MDR-ST). This led to the use of fluoroquinolones as first line antibiotic therapy options for typhoid fever, worldwide. But then, there emerged quinolone resistant strains with decreased susceptibility to ciprofloxacin⁸. Over the past two decades, global spread of a dominant, usually MDR, haplotype of *S. typhi* called H58 (genotype 4.3.1) has been observed. It emerged in early 1990s, has ability to maintain and traffic MDR plasmids and is associated with resistance to fluoroquinolones through common gyrA/parC mutations. It is prevalent across Southeast Asia and parts of Africa and Oceania. Global dispersal of this lineage is now a real possibility⁹.

In Pakistan, MDR and quinolone resistant *S. enterica* strains are of great concern for public health. As a result of emergence and spread of quinolone resistant *S. typhi* in Pakistan, 3rd generation cephalosporins such as oral cefixime or parenteral ceftriaxone became empirical treatment options¹⁰.

Since November 2016, a large proportion of ceftriaxone resistant cases have been reported in Sindh particularly Hyderabad and Karachi and these strains were XDR (extensively drug resistant) i.e. non susceptible to chloramphenicol, ampicillin, cotrimoxazole, fluoroquinolones and 3rd generation cephalosporins¹¹.

As continuous monitoring and evaluation of local antimicrobial susceptibility testing (AST) profiles are in practice to update therapeutic guidelines, the present study was done in a tertiary care hospital in Lahore, keeping in view XDR *Salmonella enterica* outbreak.

MATERIAL AND METHODS

In this prospective Study *Salmonella enterica* (*S. typhi* & *S. paratyphi* A) isolates were retrieved from blood culture samples received at Microbiology Laboratory from different departments of Shaikh

Zayed Medical Complex, Lahore during March 2018 - May 2019. A sum total of 13964 blood samples were processed for culture during study period. Data related to age, gender, serotypes of isolates and AST profiles was collected. Blood samples were first cultured in Tryptic Soy Broth followed by subculture on MacConkey and Blood agar under optimal conditions. Positive blood cultures were subjected to Identification (ID) and AST profiles using biochemical panel (API 20E) and latest CLSI recommendations, respectively. Isolates identified as *Salmonellae* were manually confirmed by slide agglutination test with polyvalent O antiserum and serovar specific antisera.

CLSI suggests following drugs to be reported for blood culture isolates of *Salmonella enterica*: ampicillin, fluoroquinolone, trimethoprim-sulfamethoxazole (cotrimoxazole), 3rd generation cephalosporin, chloramphenicol (if requested) and azithromycin. In our study, the evaluation of antibiogram was carried out as per CLSI. Azithromycin and meropenem (carbapenem) were taken into consideration as second line drugs.

Statistical analysis:

The data was computed using MS Excel and results were calculated and presented as column or row percentages.

RESULTS

Out of 13964 processed blood samples, 899 *Salmonella enterica* positive blood cultures were isolated over the span of 15 months. of these, 849 (94.4%) were identified as *S. typhi* and 50 (5.6%) as *S. paratyphi* A. (2 isolates identified as *Salmonella paratyphi* B were excluded from study due to low count). The age of the patients suffering from typhoid fever was from 2 months -67 years. Pediatric patients 81.7% accounted for majority of cases while adults were 18.3%. Age distribution is shown in Fig-1 and gender distribution in Table-1.

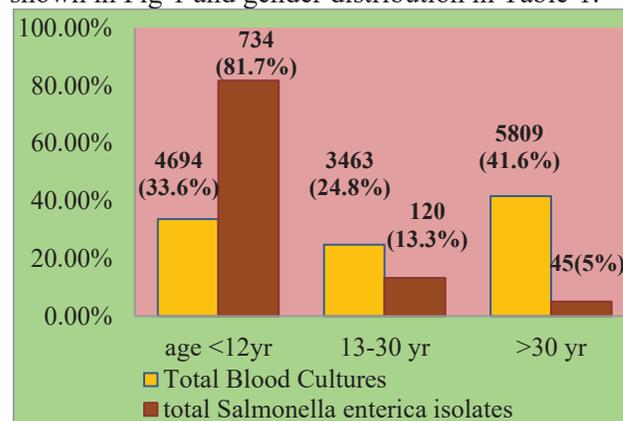


Fig-1: Age distribution

	Total Males	Total Females	Total
Total Blood Cultures	8923 (63.9%)	5041 (36.1%)	13964
Total <i>Salmonella enterica</i> isolates	513 (57.1%)	386 (42.9%)	899
Total CRO-Resistant <i>S. enterica</i> isolates	182 (35.4%)	196 (50.8%)	390

Table-1: Gender distribution

Out of 43.4% CRO-Resistant strains of XDR *S. enterica*, percentage was 44.2% and 17.6% for *S. typhi* and *S. paratyphi* A respectively. *S. enterica* isolates were 42.1% resistant to cefixime. Out of this, 43.7% resistance was observed in *S. typhi* and 11.8% in *S. paratyphi* A. Following is a graph showing total number of blood cultures received in each month of research study (Fig-2).

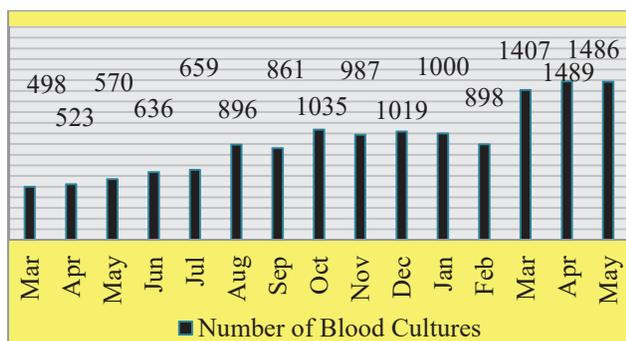


Fig-2: Number of Blood Cultures (March 2018-May 2019)

There is 2.8 times increase in the number of total blood cultures, over a span of 15 months. In addition, when three months (March, April and May) of two consecutive years (2018 & 2019) were compared, there was a significant increase of 275% in the count.

A graph showing consistently rising resistance percentage to 3rd generation cephalosporins i.e. ceftriaxone (CRO) among *S. enterica* isolates over the span of 15 months is shown in Fig-3. It shows ever increasing percentages, even per month of the ongoing outbreak.

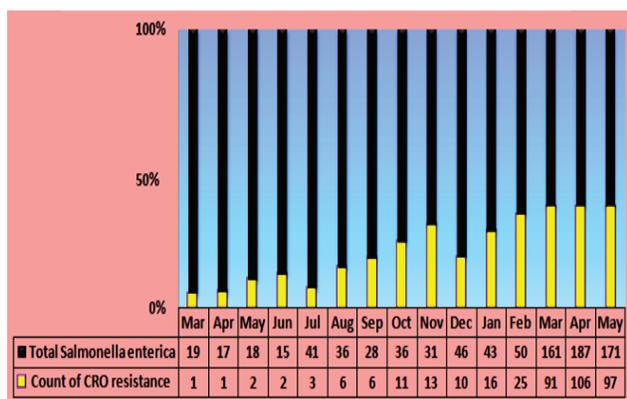


Fig-3: CRO-resistant *Salmonella enterica* isolates vs total *Salmonella enterica* isolates (March 2018 - May 2019)

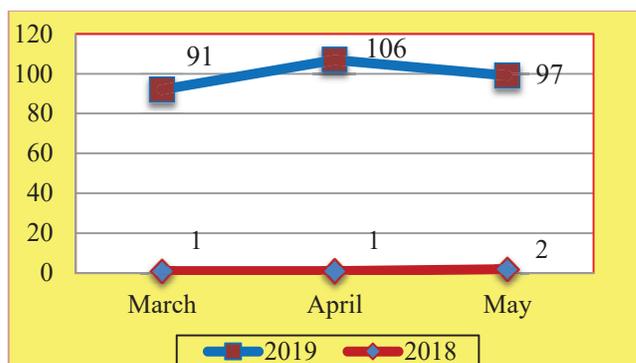


Fig-4: Comparison of CRO - Resistance (2018 vs 2019)

Fig-4 shows comparison of 3rd generation cephalosporin (ceftriaxone) resistance for three months (March, April and May) of two consecutive years 2018 and 2019. There was a marked difference in the resistance pattern of the antimicrobial agent when compared after one year. Moreover, all isolates tested for azithromycin and meropenem were susceptible to these antibiotics.

DISCUSSION

Pakistan is currently facing the largest outbreak of extensively drug resistant invasive Salmonellosis. Reports from various districts have confirmed this fact¹.

A study at Hyderabad revealed largest outbreak of CRO-resistant *S. typhi* in the history of Pakistan that started at November 2016. Investigation discovered the likely reason of fecal contamination of water supply, contact with infected individuals and irrational antibiotic use.¹¹ Our present study was aimed to monitor the ever changing landscape of *S. enterica* antimicrobial resistance. 3rd generation cephalosporin resistance of XDR *S. typhi* was the main focus. Parenteral forms of 3rd generation cephalosporins, considered as empirical therapy options in complicated typhoid fever cases before this outbreak, have equivalent efficacy when used in typhoid fever complications (e.g. meningitis) whereas cefixime qualifies as highly effective and safe oral drug with an efficacy of 92.5% in a study.^{12,13} These drugs have been proved as resistant in almost half of the cases of typhoid fever in our study. The count of total blood cultures, *S. enterica* isolates and ceftriaxone resistant *S. enterica* isolates were low in earlier months but all these parameters increased towards the end of research study as depicted in Fig-2 & 3. For investigation of enteric fever, increasing trend for blood culturing reinforced by recent infection control policies, has definitely highlighted its importance as gold standard diagnostic tool compared to different non

significant serological tests. In this way, it has yielded increased count of CRO-resistant *S. enterica* strains as well. This high cephalosporin resistance has also been reported from other areas of Asia (i.e. Bangladesh and India).¹¹ Several case reports from west have raised the same issue e.g. a Canadian report revealed first case of CRO-resistant *S. typhi* in a three year old child who visited Karachi, Pakistan and got sick.¹⁴ Many other serovars of *S. enterica* have also been implicated in cephalosporin resistance for example XDR *S. enterica* serovar Oranienburg originated in Pakistan in recent years¹⁵. Although maximum percentage of blood culture samples was retrieved from patients of more than 30 years of age in our research, the pediatric age group (<12 years) had invasive Salmonellosis at its peak (81.7%), a result consistent with many other research studies. The likely reasons considered for such higher resistance percentage are earlier consultation for treatment by the guardians of children, low trend of self-medication (antibiotic use) and increased microbial load (i.e. colony forming units/ml of blood) in children as compared to adults. Among adults, male preponderance (57.1%) was observed for typhoid fever cases, a result consistent with many other research studies e.g. an Indian research showed 58% male typhoid cases¹⁶ and a study in Bangladesh revealed 54% cases¹⁷ in contrast to an Eastern Anatolian research showing male to female ratio of 1:1.5 (61:93).¹⁸ Social and behavioral variations in different areas of world /age group affected or both might be the reason for difference.¹¹ However, XDR *Salmonella* infection cases were from female patients mainly in the current study.

All strains tested for azithromycin and meropenem were 100% susceptible to these agents, a result same as many other studies.

CONCLUSION

The in vitro susceptibility patterns of 899 *Salmonella enterica* (*S. typhi* & *S. paratyphi* A) isolates showed consistently high resistance to 3rd generation cephalosporins (parenteral as well as oral), the drugs deemed as empirical therapy options in recent past. For these typhoid fever causing XDR strains, 2nd line agents like carbapenems and azithromycin seem to be the last resort drugs. That is really an alarming situation as it in turn may have serious implications in our resource poor country due to high cost and little accessibility.

Acknowledgements:

We owe special thanks to Prof. Khalid Mehmood (Microbiology Department, College of Medicine & Dentistry University of Lahore) and Dr. Aqsa Aslam (Assistant Professor of Microbiology at Sharif Medical and Dental College, Lahore) for reviewing and editing the article.

REFERENCES

1. World Health Organization Regional Office for Eastern Mediterranean. Weekly epidemiological monitor: disease outbreaks in Eastern Mediterranean Region (EMR), January to December 2018. Cairo, Egypt: World Health Organization Regional Office for Eastern Mediterranean; 2018.
2. Mogasale V, Maskery B, Ochiai RL, et al. Burden of typhoid fever in low-income and middle income countries: a systematic literature based update with risk factor adjustment. *Lancet Globe Health*. 2014; 2:e570-80.
3. Crump JA, Luby SP, Mintz ED. The global burden of typhoid fever. *Bull world Health Organ*. 2004; 82:346-53.
4. Ochiai RL, Acosta CJ, Danovaro-Holliday MC et al. Domi Typhoid Study Group. A study of typhoid fever in five Asian countries: disease burden and implication for controls. *Bull world Health Organ*. 2008; 86: 260-68.
5. Cooke FJ, Wain J. The Emergence of Antibiotic Resistance in Typhoid Fever. *Travel Med Infect Dis*. 2004; 2: 67-74.
6. Parry C, Hien T, Dougan G, et al. Typhoid fever. *N Engl J Med*. 2002; 347: 1770-82.
7. Saxena SN, Sen R. *Salmonella paratyphi* A infection in India: incidence and phage types. *Trans R Soc Trop Med Hyg*. 1966; 60: 409-11.
8. Walia M, Gaiind R, Mehta R, Paul P, Aggarwal P, Kalaiwani M. Current perspectives of enteric fever: a hospital-based study from India: *Ann Trop Paediatr*. 2005; 25: 161-74.
9. Wong VK, Baker S, Pickard DJ, et al. Phylogeographical analysis of the dominant multidrug-resistant H58 clade of *Salmonella Typhi* identifies inter- and intracontinental transmission events. *Nat Genet*. 2015; 47:632-9.
10. Qamar FN, Azmatullah A, Kazi A, et al. A three year review of antimicrobial resistance of *Salmonella enterica* serovars Typhi and Paratyphi A in Pakistan. *J Infect Dev Ctries*. 2014; 8: 981-6.
11. Qamar FN, Yousafzai M, Khalid M, et al. Outbreak investigation of ceftriaxone-resistant *Salmonella enterica* serotype Typhi its risk

- factors among the general population in Hyderabad, Pakistan; a matched case-control study. *The Lancet Infectious Diseases*. 2018; 12: 1368-76.
12. Kavoliotis J, Tsiaousi A, Papavasiliou D, Kansouzidou A. Non-typhoid *Salmonella* meningitis. *Scandinavian journal of infectious diseases*. 1994; 26.4: 403-5.
 13. Chaudhry MK, Rayamajhi BS, Paudel K, et al. Efficacy of cefixime in the treatment of typhoid fever. *International Journal of Pharmaceutical & Biological Archives*. 2014; 4(2): 307-9.
 14. Wong W, Al Rawahi H, Patel S, Yau Y, Eshaghi A, et al. The first Canadian Pediatric case of extensively drug-resistant *Salmonella* Typhi originating from an outbreak in Pakistan and its implication for empiric antimicrobial choices. *IDCases*. 2019; 15:e00492.
 15. Yang WC, Chan OW, Wu TL, et al. Development of ceftriaxone resistance in *Salmonella enterica* serotype Oranienburg during therapy for bacteremia. *Journal of Microbiology, Immunology and Infection*. 2016; 1:41-5.
 16. Capt RMS, Capt KBS, Col SML, Col AN. Outbreak of multidrug resistant S Typhi enteric fever in Mumbai Garrison. *Med J Armed Forces India*. 2005; 61:148-50.
 17. Ram P, Naheed A, Brooks W, et al. Risk factors for typhoid fever in slum in Dhaka, Bangladesh. *Epidemiol Infect*. 2007; 135:458-65.
 18. Aypak A, Çelik AK, Aypak C, Çikman Ö. Multidrug resistant typhoid fever outbreak in Ercek Village-Van, Eastern Anatolia, Turkey: clinical profiles, sensitivity patterns and response to antimicrobials. *Trop Doct*. 2010; 40: 160-2.

The Authors:

Dr. Sadia Hameed,
P.G. Trainee,
Department of Microbiology,
Shaikh Zayed Medical Complex, Lahore.

Prof. Mateen Izhar
Head of Microbiology Department,
Shaikh Zayed Medical Complex, Lahore.

Dr. Anwaar Basheer,
Associate Professor,
Department of Microbiology,
Shaikh Zayed Medical Complex, Lahore.

Dr. Chetan Lal,
Assistant Professor,
Department of Microbiology,
Shaikh Zayed Medical Complex, Lahore.

Dr. Shazia Rishi,
Senior Medical Officer,
Department of Microbiology,
Shaikh Zayed Medical Complex, Lahore.

Abdul Basit,
Laboratory Manager,
Department of Microbiology,
Shaikh Zayed Medical Complex, Lahore.

Corresponding Author:

Dr. Sadia Hameed,
P.G. Trainee,
Department of Microbiology,
Shaikh Zayed Medical Complex, Lahore.
E-mail: sadiahameed888@gmail.com